

2,5-Dichloroaniline, a monoclinic structure with a pseudo-tetragonal unit cell

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Key indicators

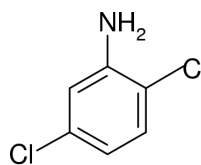
Single-crystal X-ray study
 $T = 120$ K
Mean $\sigma(\text{C}-\text{C}) = 0.002$ Å
 R factor = 0.026
 wR factor = 0.074
Data-to-parameter ratio = 10.7For details of how these key indicators were
automatically derived from the article, see
<http://journals.iucr.org/e>.

The pseudo-tetragonal cell of the title compound, $\text{C}_6\text{H}_5\text{Cl}_2\text{N}$, is correctly described as monoclinic with $\beta = 90.033(2)^\circ$. Amine groups are linked by intermolecular hydrogen bonding involving only one H atom of each group.

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Comment

The crystal structure of 2,5-dichloroaniline, (I), was previously determined (Sakurai *et al.*, 1963) from Weissenberg film data. The space group reported was $P2_1/c$ and refinement converged at $R = 0.126$. The original cell dimensions were $a = 13.237(7)$, $b = 3.892(6)$, $c = 18.80(2)$ Å and $\beta = 135.2(2)^\circ$. The large β angle led to the examination of the cell dimensions with *LEPAGE* (Spek, 1988) and the possibility of a tetragonal or orthorhombic cell was suggested. A new data set collected at 120 K refined to $R = 0.026$ in the space group $P2_1/n$ with cell dimensions $a = 13.1141(7)$, $b = 3.8137(6)$, $c = 13.1699(7)$ Å and $\beta = 90.033(2)^\circ$. Final analysis with *PLATON* (Spek, 2001) showed that the lattice featured metrical symmetry (pseudo-tetragonal or pseudo-orthorhombic) not supported by the cell contents which confirmed the crystal system as monoclinic. The $P2_1/n$ designation is related to the size of the β angle, which is much closer to 90° in this setting, compared to the transformed cell. Taking the temperature of the determination into consideration, the cell dimensions originally reported and the cell dimensions transformed into $P2_1/c$ from this study are equivalent; hence polymorphism is not shown.



(I)

Details of the 2,5-dichloroaniline structure (Fig. 1) not previously reported include hydrogen-bond formation between amine groups that involves only one of the H atoms (H1B) (Fig. 2). This results in continuous chains of molecules running in the direction of the b axis that each contain two of the four symmetry-related molecules per unit cell required by the space group (Fig. 3). The aromatic rings pack face-to-face to each other in these chains by translational symmetry along the b axis. The close separation of these rings (3.490 Å) indicates π - π -stacking interactions. There is also a short intramolecular H1B...Cl1 separation of 2.63(2) Å but the N1—H1B...Cl1 angle is only $107(2)^\circ$. The N atom deviates by 0.24(1) Å from the plane defined by atoms C1, H1A and H1B.

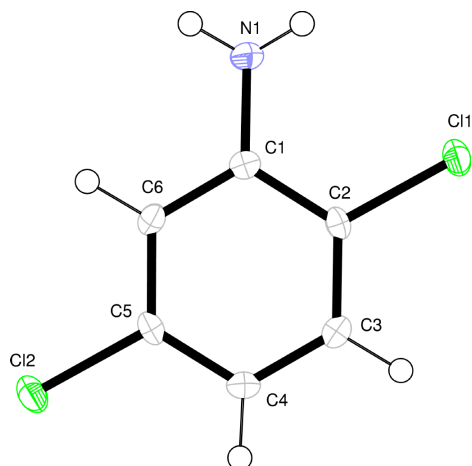


Figure 1
The molecular structure of (I). Displacement ellipsoids are shown at the 50% probability level.

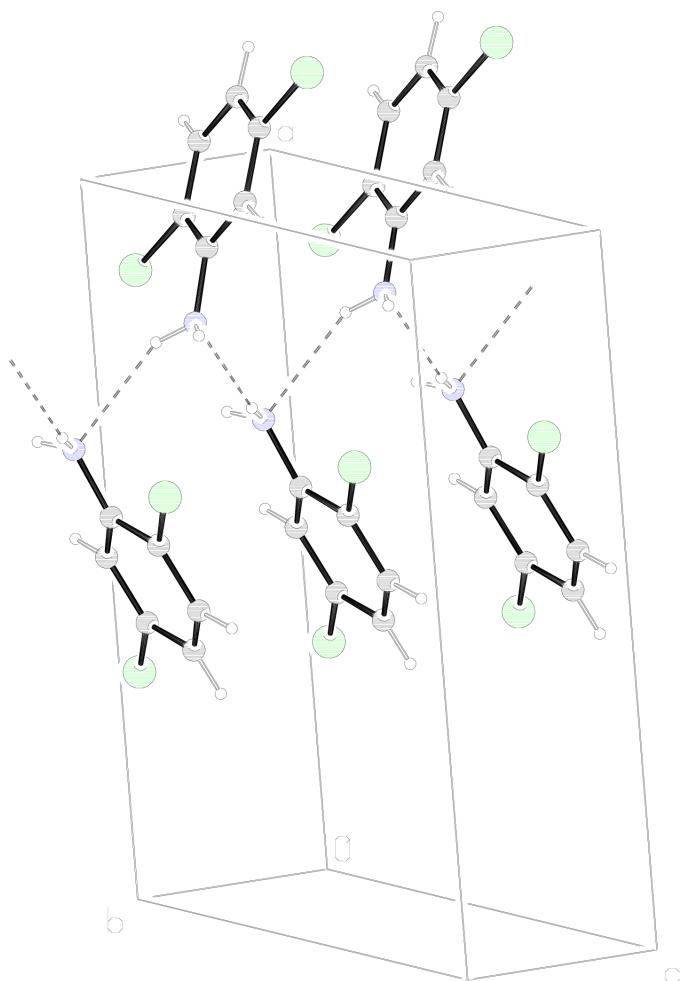


Figure 2
Crystal packing diagram showing (x, y, z) molecules linked to $(-x + \frac{3}{2}, y + \frac{1}{2}, -z + \frac{1}{2})$ molecules by translation along the b axis.

The shortest $\text{Cl}\cdots\text{Cl}$ separation is $\text{Cl1}\cdots\text{Cl2}(-\frac{1}{2} + x, \frac{1}{2} - y, -\frac{1}{2} + z) = 3.3219$ (8) Å, compared to the previously reported value of 3.37 Å.

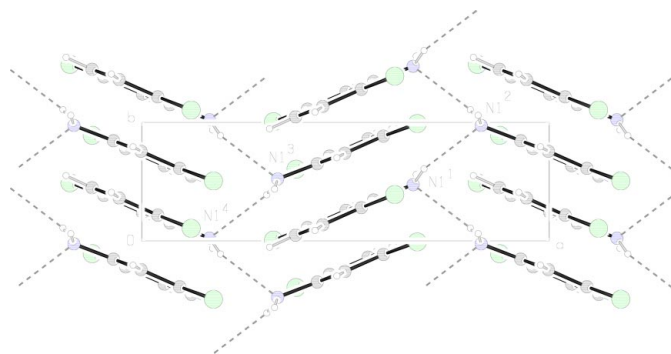


Figure 3
Crystal packing diagram showing the two chains of hydrogen-bonded molecules within the unit cell. [Symmetry-code suffixes: (1) x, y, z ; (2) $\frac{3}{2} - x, \frac{1}{2} + y, \frac{1}{2} - z$; (3) $1 - x, 1 - y, 1 - z$; (4) $-\frac{1}{2} + x, \frac{1}{2} - y, \frac{1}{2} + z$.]

The crystal structures of 2,3-, 2,4-, 2,6-, 3,4- and 3,5-dichloroaniline (Dou *et al.*, 1993) also show $\text{N}-\text{H}\cdots\text{N}$ hydrogen bonds and short $\text{Cl}\cdots\text{Cl}$ interactions.

Experimental

2,5-Dichloroaniline was purchased from Aldrich and colourless crystals were obtained by sublimation.

Crystal data

$\text{C}_6\text{H}_5\text{Cl}_2\text{N}$	$D_x = 1.634 \text{ Mg m}^{-3}$
$M_r = 162.01$	Mo $K\alpha$ radiation
Monoclinic, $P2_1/n$	Cell parameters from 1724 reflections
$a = 13.1141$ (6) Å	$\theta = 2.9\text{--}26.0^\circ$
$b = 3.8137$ (1) Å	$\mu = 0.88 \text{ mm}^{-1}$
$c = 13.1699$ (7) Å	$T = 120$ (2) K
$\beta = 90.033$ (2)°	Lozenge, colourless
$V = 658.67$ (11) Å ³	$0.20 \times 0.15 \times 0.10 \text{ mm}$
$Z = 4$	

Data collection

Enraf-Nonius KappaCCD area-detector diffractometer	1088 independent reflections
φ and ω scans to fill Ewald sphere	1026 reflections with $I > 2\sigma(I)$
Absorption correction: multi-scan (SORTAV; Blessing, 1995)	$R_{\text{int}} = 0.033$
$T_{\text{min}} = 0.844, T_{\text{max}} = 0.917$	$\theta_{\text{max}} = 26.0^\circ$
2663 measured reflections	$h = -16 \rightarrow 16$
	$k = -4 \rightarrow 4$
	$l = -16 \rightarrow 11$

Refinement

Refinement on F^2	$w = 1/[\sigma^2(F_o^2) + (0.0449P)^2 + 0.1577P]$
$R[F^2 > 2\sigma(F^2)] = 0.026$	where $P = (F_o^2 + 2F_c^2)/3$
$wR(F^2) = 0.074$	$(\Delta/\sigma)_{\text{max}} = 0.001$
$S = 1.03$	$\Delta\rho_{\text{max}} = 0.26 \text{ e \AA}^{-3}$
1088 reflections	$\Delta\rho_{\text{min}} = -0.36 \text{ e \AA}^{-3}$
102 parameters	
All H-atom parameters refined	

Table 1

Selected geometric parameters (Å, °).

$\text{Cl1}-\text{C2}$	1.7356 (17)	$\text{Cl2}-\text{C5}$	1.7432 (18)
$\text{C6}-\text{C1}-\text{C2}$	117.46 (13)	$\text{C3}-\text{C4}-\text{C5}$	118.24 (14)
$\text{C3}-\text{C2}-\text{C1}$	121.36 (15)	$\text{C4}-\text{C5}-\text{C6}$	121.61 (16)
$\text{N1}-\text{C1}-\text{C2}-\text{Cl1}$	4.6 (2)		

Table 2
Hydrogen-bonding geometry (Å, °).

$D-H \cdots A$	$D-H$	$H \cdots A$	$D \cdots A$	$D-H \cdots A$
$N1-H1B \cdots N1^i$	0.87 (2)	2.45 (2)	3.241 (2)	151 (2)

Symmetry code: (i) $\frac{3}{2} - x, \frac{1}{2} + y, \frac{1}{2} - z$.

Refined N–H and C–H distances are in the ranges 0.85 (2)–0.87 (2) and 0.93 (2)–0.97 (2) Å, respectively.

Data collection: *DENZO* (Otwinowski & Minor, 1997) and *COLLECT* (Hooft, 1998); cell refinement: *DENZO* and *COLLECT*; data reduction: *DENZO* and *COLLECT*; program(s) used to solve structure: *SIR97* (Altomare *et al.*, 1999); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *ORTEP-3* (Farrugia, 1997) and *PLATON* (Spek, 2001); software used to prepare material for publication: *SHELXL97* and *PLATON*.

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References

- Altomare, A., Burla, M. C., Camalli, M., Cascarano, G. L., Giacovazzo, C., Guagliardi, A., Moliterni, A. G. G., Polidori, G. & Spagna, R. (1999). *J. Appl. Cryst.* **32**, 115–119.
- Blessing, R. H. (1995). *Acta Cryst.* **A51**, 33–38.
- Dou, S., Weiden, N. & Weiss, A. (1993). *Acta Chim. Hung.* **130**, 497–522.
- Farrugia, L. J. (1997). *J. Appl. Cryst.* **30**, 565.
- Fletcher, D. A., McMeeking, R. F. & Parkin, D. (1996). *J. Chem. Inf. Comput. Sci.* **36**, 746–749.
- Hooft, R. (1998). *COLLECT*. Nonius BV, Delft, The Netherlands.
- Otwinowski, Z. & Minor, W. (1997). *Methods in Enzymology*, Vol. 276, *Macromolecular Crystallography*, Part A, edited by C. W. Carter & R. M. Sweet, pp. 307–326. London: Academic Press.
- Sakurai, T., Sundaralingam, M. & Jeffrey, G. A. (1963). *Acta Cryst.* **16**, 354–363.
- Sheldrick, G. M. (1997). *SHELXL97*. University of Göttingen, Germany.
- Spek, A. L. (1988). *J. Appl. Cryst.* **21**, 578–579.
- Spek, A. L. (2001). *PLATON*. Utrecht University, The Netherlands.